



AXYS Analytical Services Ltd.

**“EXPERIENCES in ANALYSIS of PPCPs
and STEROIDS/HORMONES in POTW
BIOSOLIDS”**

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NELAP ACCREDITED

ISO 17025 CERTIFIED



AGENDA

- ❑ **Overview of Analytical Methods**
 - ❑ **PPCP Method**
 - ❑ **Hormone / Sterol Method**
- ❑ **Analytical Challenges**
 - ❑ **Clean Matrices vs. POTW Matrices**
 - ❑ **Positive Identification / False Positives and Suppression**
 - ❑ **Range of Analyte Concentration**
 - ❑ **Sample Handling / Preservation**



PURPOSE of ANALYSIS

- ❑ **Ultra- Trace DL Targets (pg to ng)**
- ❑ **PPCP Method**
 - ❑ **Tool to Investigate Levels and Occurrence of PPCPs in Environment**
 - ❑ **73 Targets**
- ❑ **HORMONE / STEROL Analysis**
 - ❑ **Tool to Investigate Levels and Occurrence of Hormones and Sterols in Environment**
 - ❑ **27 Targets**
 - ❑ **Source Info / Fingerprinting**



COMMON to BOTH ANALYSIS

- ❑ **Developed as per EPA 1600 Series Methods**
 - ❑ Performance based
 - ❑ Multi Matrix purpose and validation
 - ❑ Validated by Federal Register 40 CFR, Part 136, Appendix B
- ❑ **Utilize labeled surrogate addition at start of process, recovery corrected vs. surrogates**
- ❑ **Validated for Positive Identification as part of large analyte panel**
- ❑ **Focus on native compounds, not degradation products or metabolites**



Rationale for Analyte Selection

- ❑ **Current or Historic Use / Occurrence**
- ❑ **Availability of Standards and Appropriate Surrogates**
- ❑ **Capture by SPE Cartridge (PPCP)**
- ❑ **Performance in Derivatization Process (Hormones and Sterols)**
- ❑ **Analyte Stability (Matrix and Instrument)**
- ❑ **Appropriate Monitoring Ions**



BATCH QC

- Max. Batch Size (including QC) is 20 samples (8 to 14 is more common)
- Each Batch contains:
 - method blank
 - duplicate
 - Bracketing Cal.
 - Instrument Blank
 - MS/MSD, SRM, CRM included by request and applicability



OVERVIEW of PPCP ANALYSIS in POTW BIOSOLIDS

Sample – 2 X 1 gram wet solids
Acidic Fraction to pH 2 + EDTA stabilizer, Basic Fraction to pH10

Respective Fractions – Add labeled Surrogates
Acidic Fraction – 3X ultrasonic extraction with phosphate buffer/ACN
Basic Fraction – 3x ultrasonic extraction with H₂O/NH₄OH

SPE Cartridge Cleanup (1 gram Waters Oasis HLB)
SPE Steps – condition, load, rinse, elute, reduce, transfer to MeOH
Add recovery standards

ANALYSIS by LC/MS/MS (x4)
ACIDIC EXTRACT – Neg. Ionization, Tetracyclines, Pos. Ionization
BASIC EXTRACT – Pos. Ionization

PPCP Standards and Surrogates

- Non-labeled standards for each target compound
- Stable isotope labeled standards where ever possible used as internal standards
 - 17 labeled standards (^{13}C , ^2D or ^{15}N) for acidic ESI+compounds
 - Labeled (^{13}C) thiabendazole and sulfamethazine for tetracyclines
 - Exact labeled analogue for all 6 Acidic ESI - compounds
 - Labeled (^2D) albuterol and metformin for basic compounds
- Labeled injection standards (^{13}C Atrazine or 2,4,5-TPP) to quantify recovery of internal standards

PPCP QA ACCEPTANCE CRITERIA

- EPA Tier 1 Validation Protocols for Single Laboratory Method for IPR, MS/MSD, and MDL criteria
- Duplicate Criteria – 40% RPD for Analytes 10X above MDL
- Bracketing Cal. – 40% RSD for targets detected
- Cal. Ver – 25% RSD for native analytes with exact labeled standards, 35% for others
- Linearity (ICAL) – 25% RSD for native analytes that have exact labeled standards, 35% for others

PPCP Analyte List

ESI – LC/MS/MS

Acidic Extraction / + ve Ionization

ACETOMINOPHEN	CLAXACILLIN
AMPICILLIN	CODEINE
AZITHROMYCIN	COTININE
CAFFEINE	DEHYDRONIFEDIPINE
CARBADOX	DEPHENIHYRDAMINE
CARBAMAZEPINE	DILTIAZEM
CEFOTAXINE	DIGOXIN
CIPROFLAXIN	DIGOXIGENIN

PPCP Analyte List

ESI – LC/MS/MS

Acidic Extraction / + ve Ionization (cont.)

ENROFLAXIN	NORGESTIMATE
ERYTHROMYCIN – H2O	OFLOXACIN
FLUMEQUINE	ORMETOPRIM
FLUOXETINE	OXACILLIN
LINCOMYCIN	OXOLINIC ACID
LOMOFLOXACIN	PENICILLIN G
MICONAZOLE	PENICILLIN V
NORFLOXACIN	ROXITHROMYCIN

PPCP Analyte List

ESI – LC/MS/MS

Acidic Extraction / + ve Ionization (cont.)

SARFLOXACIN	TRIMETHOPRIM
SULFACHLOROPYRIDAZINE	TYLOSIN
SULFADIAZINE	VIRGINIAMYCIN
SULFADIMETHOXINE	1,7 DIMETHYLXANTHINE
SULFAMERIZINE	
SULFAMETHAZINE	
SULFAMETHAZOLE	
THIABENDAZOLE	

PPCP Analyte List

“Tetracyclines” by ESI - LC/MS/MS

Acidic Extraction / - ve Ionization

Chlortetracycline	Doxycycline
4–Epichlorotetracycline	Oxytetracycline
Anhydrochlortetracycline	4 – Epitetracycline
4-Epianhydrochlor-tetracycline	4 - Epianhydrotetracycline
Isochlortetracycline	Anydrotetracycline
Democlocycline	Mincycline

PPCP Analytes by ESI – LC/MS/MS

ACIDIC EXTRACTION / -VE IONIZATION MODE	BASIC EXTRACTION / + VE IONIZATION MODE
WARFARIN	ALBUTEROL
IBUPROFEN	CIMETIDINE
GEMFIBROZIL	METFORMIN
NAPROXEN	RANITIDINE
TRICLOSAN	
TRICLOCARBAN	

OVERVIEW of HORMONES/STEROLS ANALYSIS in POTW BIOSOLIDS

SAMPLE SIZE = 1 GRAM WET
(after discarding “debris”)

Surrogate Addition, dry with Sodium Sulfate
Soxhlet 18 hrs. 40:60 Hexane:Acetone
DCM rinses added to extract, reduce

Solvent Exchange to 5% Toluene in Hexane
Clean-Up – Layered Alumina / Florisil Column - Concentrate
Derivatize with 99:1 BSTFA;TMCS Reagent to produce Trimethylsilyl Ethers

ANALYSIS by HRGC / HRMS
or GC/MS

HORMONES / STEROLS QA ACCEPTANCE CRITERIA

- EPA Tier 1 Validation Protocols for Single Laboratory Method for IPR, MS/MSD, and MDL criteria
- Duplicate Criteria – 40% RPD for Analytes 10X above MDL
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Hormone Targets (*= dx surrogate)

ANDROSTENIDIENE	ESTRIOL
ANDROSTERONE	ESTRONE
COPROSTANOL	17 α ETHINYL ESTRADIOL *
DESOGESTRAL	MESTRANOL *
17 α DIHYDROEQUILIN	NORETHINDIONE *
EQUILENIN	NORGESTREL *
EQUILIN	PROGESTERONE *
17 α ESTRADIOL	TESTOSTERONE
17 β ESTRADIOL *	

Sterol Targets (*= dx surrogate)

CAMPESTEROL	β – SITOSTEROL
CHOLESTEROL *	β - STIGMASTRANOL
DESMOSTEROL	STIGMASTEROL
EPICOPROSTANOL	
ERGOSTEROL	
β – ESTRADIOL – 3 - BENZOATE	

BIOSOLID DETECTION LIMITS

- PPCP in validation matrix (Peat Moss)
 - Acidic Extraction / + ve Ionization (0.1 to 50 ng/g)
 - Acidic Extraction / - ve ionization (1 to 98 ng/g)
 - Tetracyclines (2-23 ng/g)
 - Basic extraction / + ve ionization (1-56 ng/g)
 - Matrix may raise DLs 10X in biosolids, target specific
- Hormones and Sterols (10 – 2000 pg/g)
 - Hormones low, sterols high
 - LR GC/MS 5-50X higher



PPCPs - CLEAN MATRICES vs. POTW MATRICES

- ❑ Removal of Matrix Interference is Limiting DL and Method QC factor , not instruments
- ❑ Some Options in Removal of Matrix Interference
 - ❑ Limit sample size
 - ❑ Wet weight preferred as POTW Biosolids Solids up to 95% H₂O with sig. matrix
 - ❑ SPE cartridge must capture targets and surrogates, excess matrix will compete
 - ❑ Increase run times, less polar mobile phases
- ❑ Representative Matrices for Validation not Available
 - ❑ Forces clean matrices DL, IPR validation (peat, sand)
 - ❑ Experience from Method QC required



HORMONES and STEROLS - CLEAN MATRICES vs. POTW MATRICES

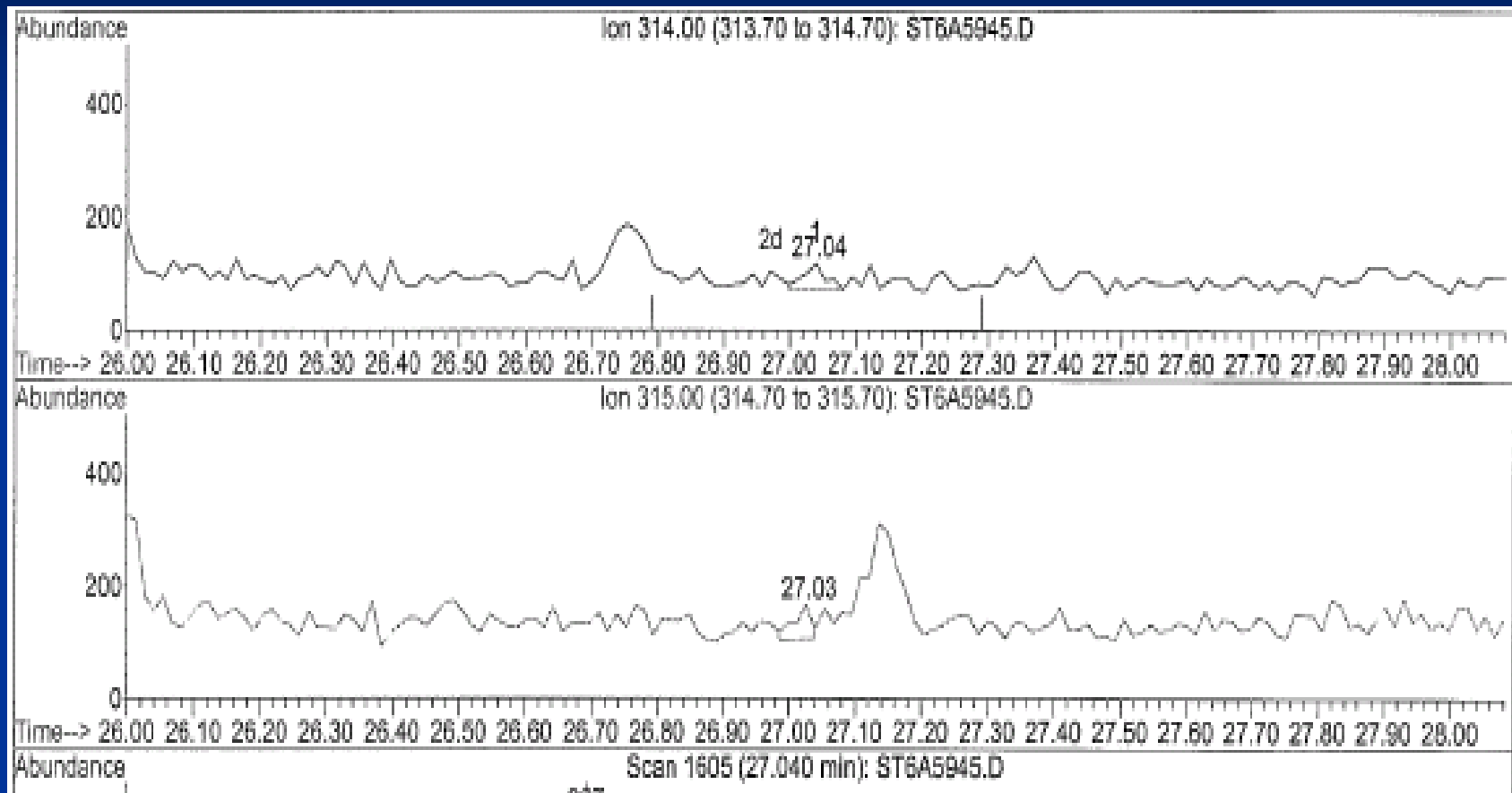
- ❑ **Removal of matrix interference is limiting DL and Method QC factor**

- ❑ **Some Options in Removal of Matrix Interference**
 - ❑ **HRMS option to improve selectivity**
 - ❑ **Can be accomplished with LC/MS/MS if high level sterols not targeted with low level hormones**
 - ❑ **Water removal vital**
 - ❑ **Limited clean-up options for large target list**

- ❑ **Representative Matrices for Validation not Available**
 - ❑ **Clean Matrices used for DL, IPR validation (peat, sand)**
 - ❑ **Experience from method QC required**



17 β Estradiol in Biosolids – GC/MS



- Native 17 β Estradiol – “NQ”

PPCPs – Suppression and Enhancement

- ❑ **Key effect in PPCP analysis by LC/MS/MS**
 - ❑ Wide variety in POTW biosolid chemistry / process
 - ❑ Varies widely by target
- ❑ **Controls**
 - ❑ Limit sample size
 - ❑ Increase capacity of clean-up (SPE type and size)
- ❑ **Monitoring and Correcting**
 - ❑ Recovery Standards to Monitor Surrogate Response
 - ❑ Dilution and re-injection when suppression or enhancement suspected



PPCPs – SUPPRESSION and ENHANCEMENT

	Biosolid A		Biosolid B		Biosolid C	
Sample size	1.14 g	3.09 g	1.16 g	3.07 g	1.11 g	3.12 g
	ng/g	ng/g	ng/g	ng/g	ng/g	ng/g
Albuterol	0.2	0.1	0.2		0.4	
Cimetidine	11.9	10.3	34.1	29.6	0.2	0.3
Metformin		10.1	40.4	21.3	28.0	15.9
Ranitidine	1.2	1.1	3.6	3.0		
d3-Albuterol	95.6	78.3	85.9	67.5	119.0	125.6
d3-Cotinine	64.3	61.9	85.5	74.2	55.5	33.3
d6-Metformin	136.9	169.5	108.8	110.7	251.8	349.4

HORMONES and STEROLS

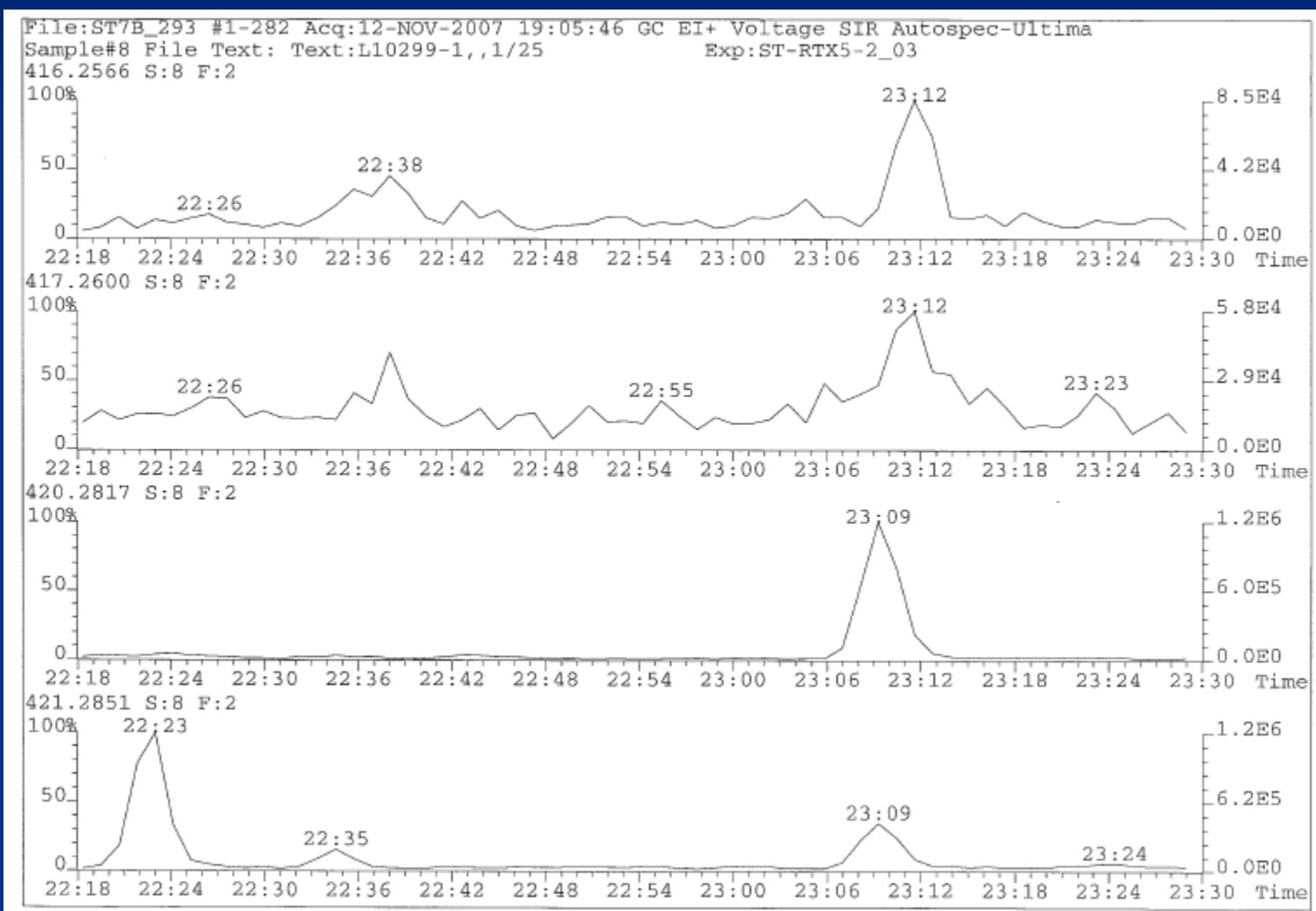
Avoiding False Positives / Negatives

- ❑ Use of internal labeled standards and relative recovery correction
- ❑ Blank acceptance criteria from Method QC (3X standard deviation, no blank correction)
- ❑ Monitor completeness of derivatization process natives and TMS products
- ❑ Increase resolution to reduce matrix effects
 - ❑ RP of GC/MS in method 100-400
 - ❑ RP of HRMS 5000 min.



17 β ESTRADIOL by HRMS

Primary / Confirming Channels



PPCPs – Concentration Range of Analytes

- ❑ High Levels produce need for dilution
- ❑ Create difficulties in MS/MSD for high level analytes
- ❑ Frequent detects above 1000 ng/g in Biosolids
 - ❑ Ciproflaxin, Ofloxacin, Triclocarban, Triclosin, ETC, Doxycycline
- ❑ Frequent detects above 1000 ng/L in POTW influent
 - ❑ Acetaminophen, Caffeine, Ibuprofen, Cotinine, Trimethoprim, Carbamazepine, Metformin, Sulfamethoxazole



HORMONES and STEROLS – Concentration Range of Analytes

- ❑ **Hormones low level, many sterols very high level in POTW biosolids**
- ❑ **Affects blank acceptance criteria**
 - ❑ **5-10 ng for most compounds**
 - ❑ **500 for cholesterol and beta sitosterol**
- ❑ **Adjustments required to prevent “NQ” on high level sterols (cholesterol and beta sitosterol most frequent)**
- ❑ **Extract split used to maintain QC**
 - ❑ **Over spike with Sterol surrogates at beginning of process**
 - ❑ **1/20 split after clean-up**
 - ❑ **2 instrument runs**
 - ❑ **Method QC adjusted to match**



SAMPLE HANDLING / PRESERVATION

- Freeze biosolid samples immediately
 - Arrest Microbiological activity
 - No definitive studies indicating hold times
 - Glass containers (PPCP and Hormones/Sterols)
- Extract immediately on thawing
- POTW processes homogenize sample
 - Allows minimization of sample size
 - Sampling plan to determine repeatability
- Have a strategy for “free water”
 - Discard, 2 analysis, attempt to re-homogenize



CONCLUSIONS

- Viable low level detection methods available for large panels of PPCPs and Hormones / Sterols
- Basic methods applicable for environmental levels (water, soil, and sediment) and POTW matrices (Influent, Effluent, Biosolids)
- High variability in POTW sludge matrix effects and levels, much work required to characterize vs. specific attributes
- High variability requires robust methods and “coping” strategies



NEXT STEPS

- Current - Peer Review of method performed by EPA – Q3/4 2007
- Further processing of samples to develop further statistical information
- Detailed Information targeted for publication by AXYS clients
- Multi-Lab Validation?
- Metabolites and Degradation Products
- Questions?



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